

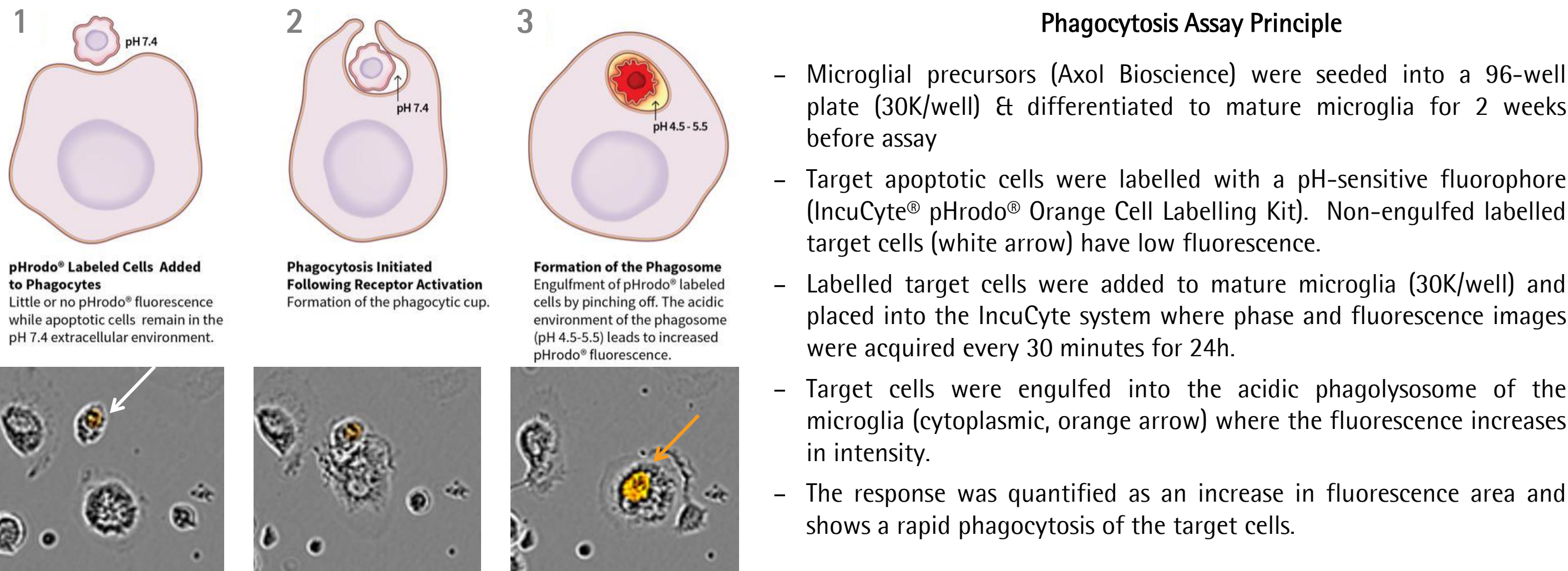
Live-cell analysis of uptake of neuropathology-associated peptides by human iPSC-derived microglia

G. Lovell, M. Bowe, S. Lopez Alcantara, J. Rauch, L. Oupicka, A. Overland, C. Schramm, T. Dale, D. Trezise
Essen BioScience, Welwyn Garden City, AL7 3AX UK & Ann Arbor, MI, 48108, USA

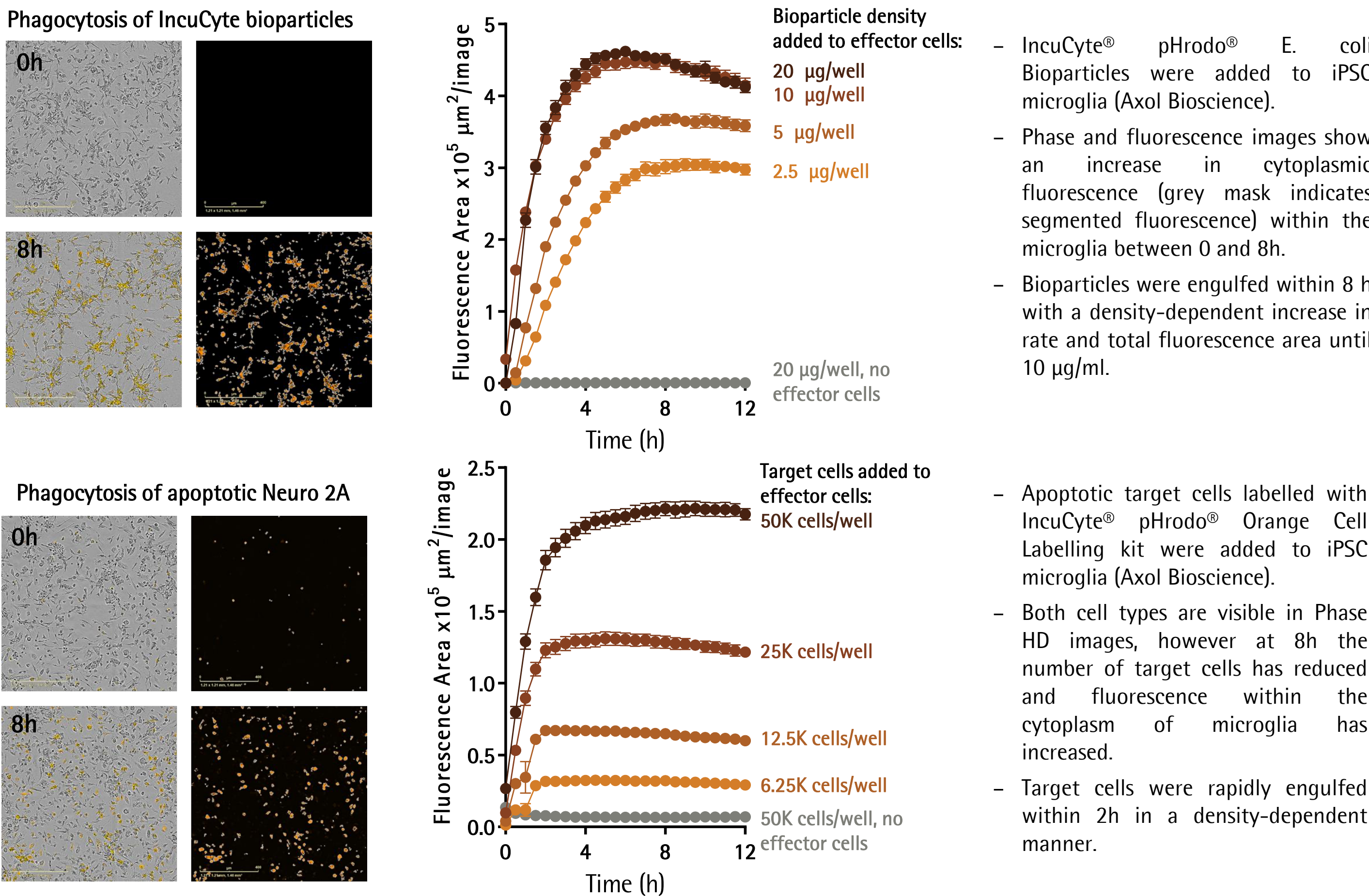
Summary & Impact

- The presence of aggregated peptides such as Amyloid β (1-42) and α -Synuclein are associated with disease phenotypes (Alzheimer's disease and Parkinson's disease, respectively).
- Microglia, as the resident macrophage of the brain, have several functions including clearance of peptide aggregates (phagocytosis) and apoptotic cells (efferocytosis).
- Here we describe characterisation of the activation and function of microglia using IncuCyte® live-cell analysis.
- Phase and fluorescence images were captured with IncuCyte® and segmented fluorescence was quantified.
- In all cases robust, time-dependent signal changes were observed, consistent with known microglia function.
- We conclude that live-cell analysis is a flexible and powerful method for analysing microglia activity, where morphological and functional parameters can be readily quantified and integrated over time.

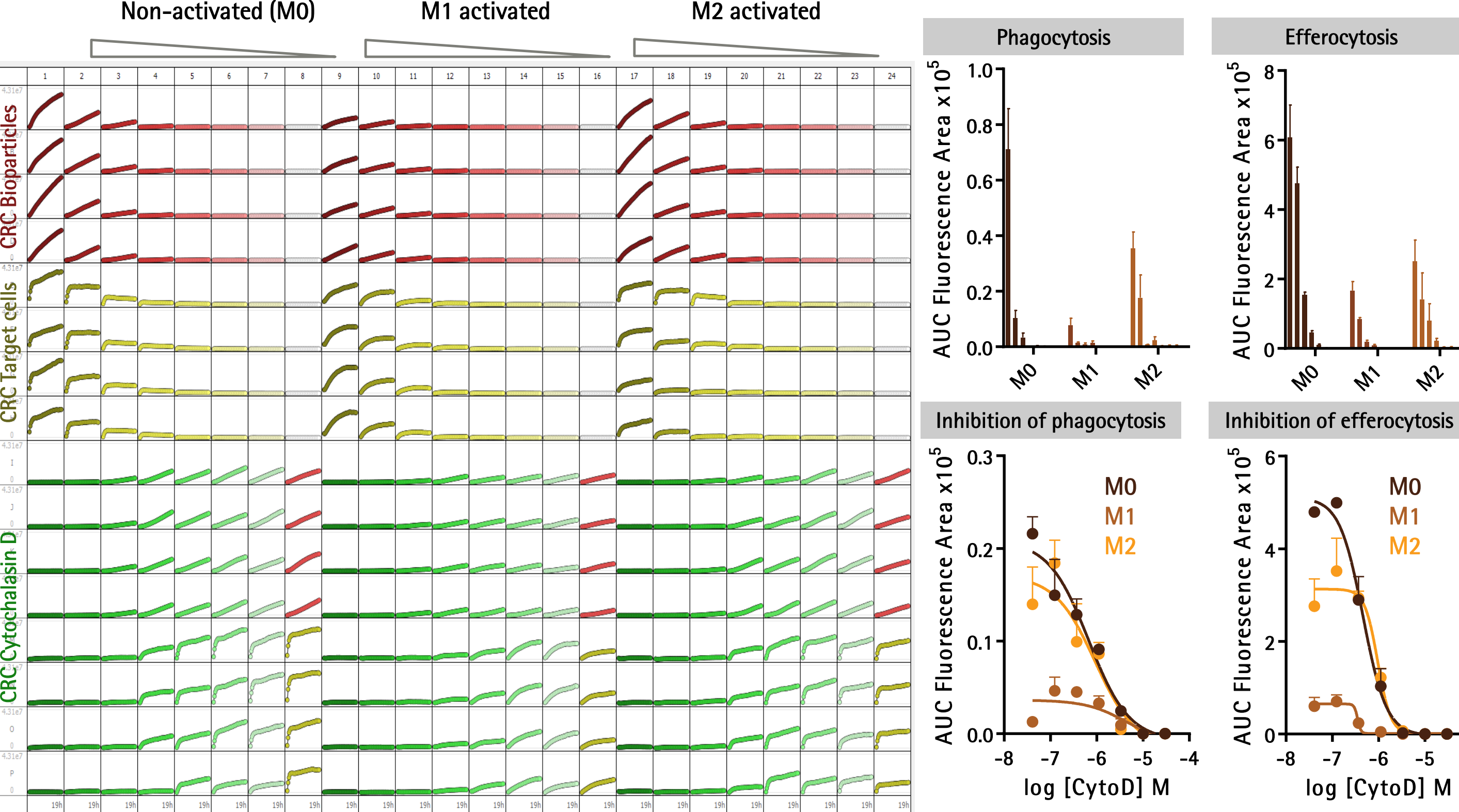
Kinetic quantification of phagocytosis by Microglia



Live-cell analysis and quantification of Microglia function

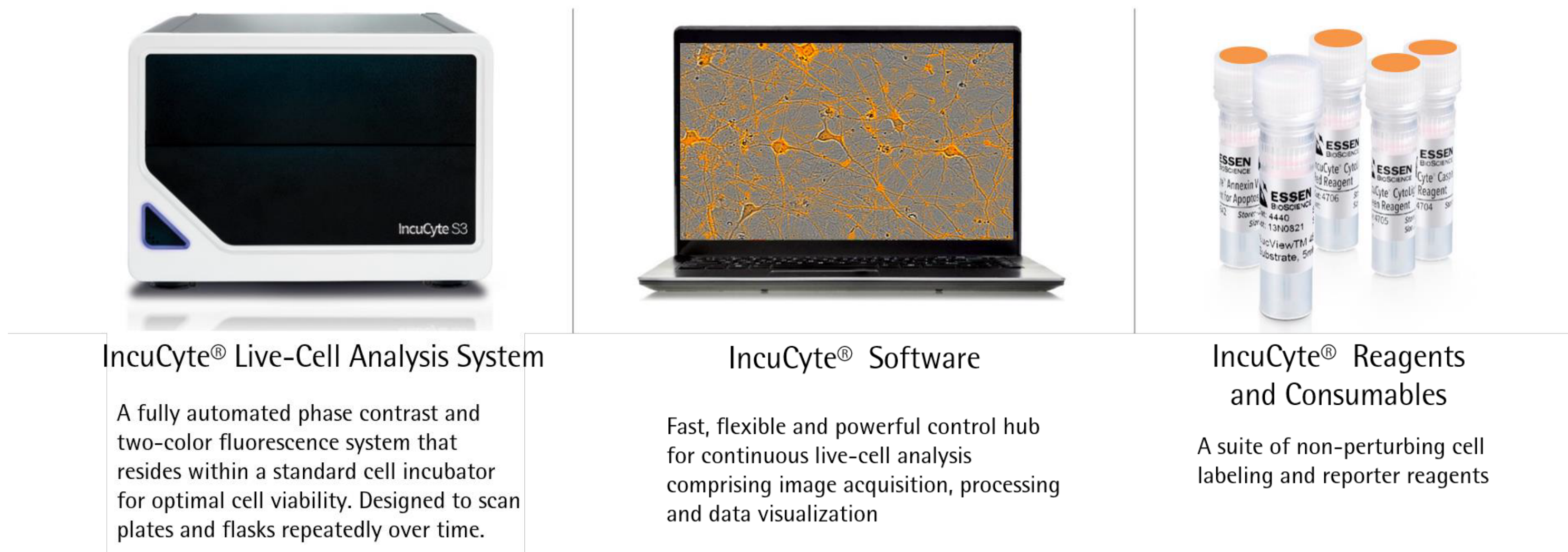


Activation state of Microglia affects phagocytic function

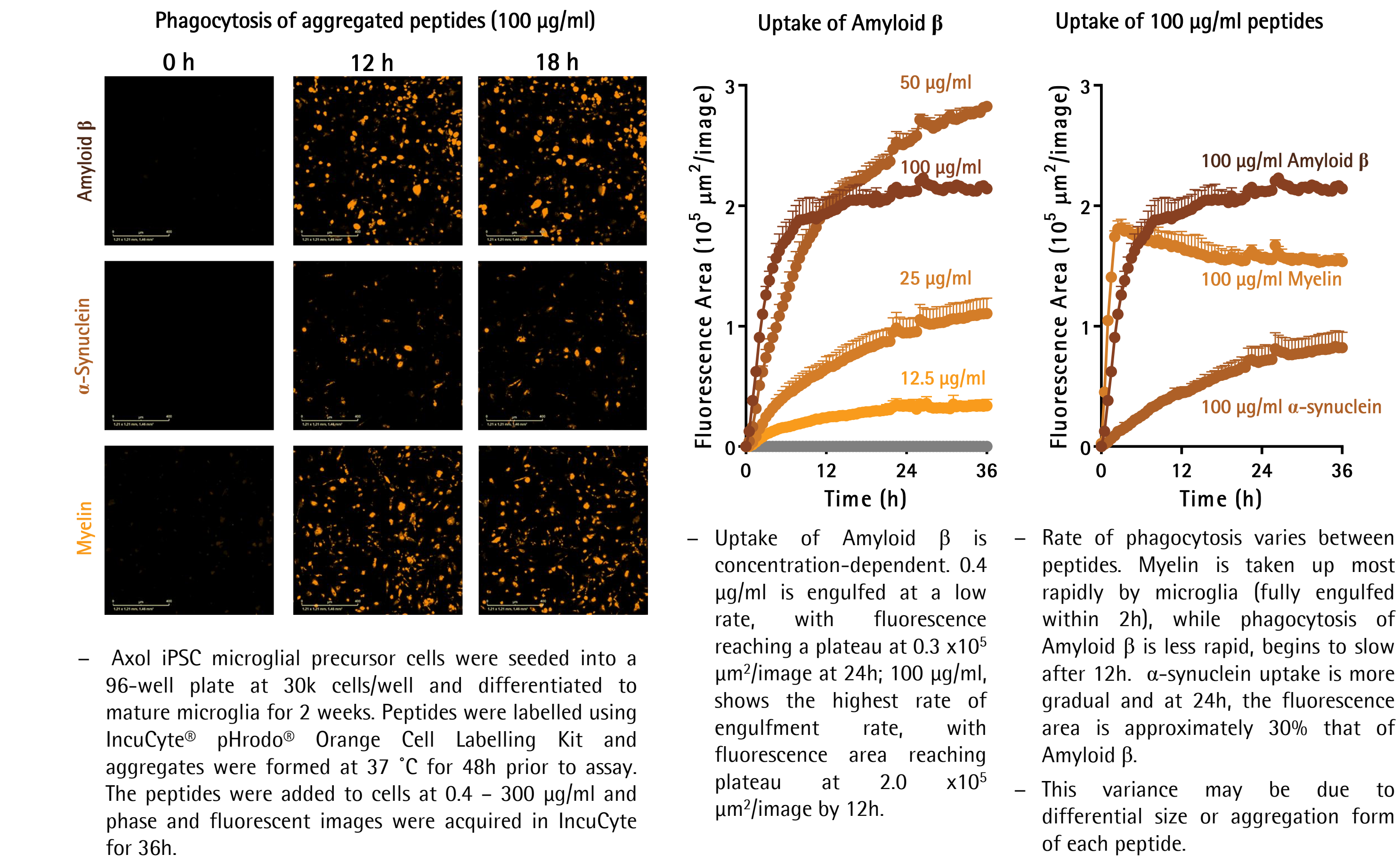


- iPSC Microglia (Axol Bioscience) were left unactivated (M0, col 1-8) or treated with LPS and IFN γ (M1, col 9-16) or IL-4 and IL-13 (M2, col 17-24) to induce polarisation of the cells. Half of these cells (rows I – P) were then treated with a concentration range of Cytochalasin D, an inhibitor of phagocytosis.
- Target material (IncuCyte® pHrodo® E. coli Bioparticles or apoptotic Neuro2A labelled with IncuCyte® pHrodo® Orange Cell Labelling Kit) was added at increasing concentrations (rows A-H) or at a single density (rows I-P).
- The plate view shows an overview of phagocytosis, plotting fluorescence area over time per well of the 384-well plate.
- In both cases microglia activated to M1 phenotype had the lowest phagocytic function with unactivated cells (M0) showing the highest rate of both phagocytosis and efferocytosis. CytoD inhibits both phagocytosis and efferocytosis in a concentration dependent manner.

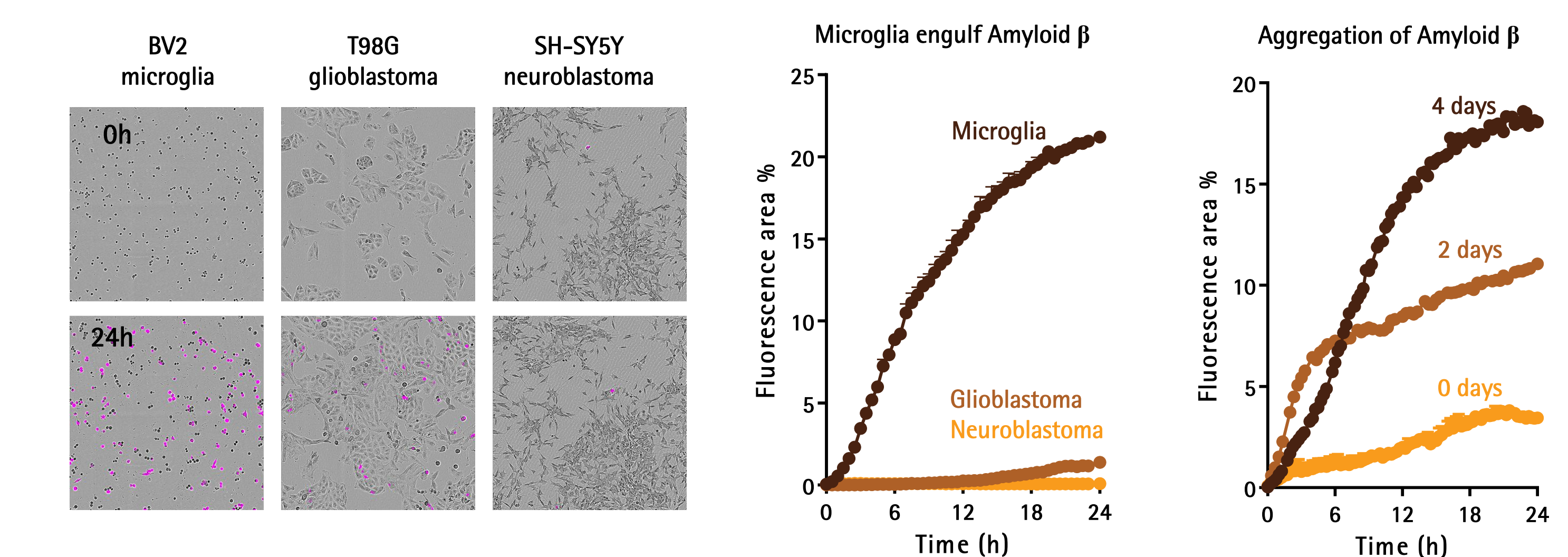
IncuCyte® System for continuous live-cell analysis: Methodology



iPSC Microglia engulf aggregated Amyloid β , Myelin and α -synuclein



Engulfment of A β is dependent on cell type and peptide aggregation



- Internalisation of aggregated labelled Amyloid β by three cell lines was compared. Phase HD images show differences in morphology between cell lines and the increase in fluorescence area (pink mask) at 24h. Data indicates the change in normalised fluorescence area over time (fluorescence area was normalised to phase area per well to account for proliferation and variations in cell morphology).
- BV-2 microglia cells are phagocytic and rapidly engulf aggregated peptide with concomitant increase in fluorescence area over 24h, while glioblastoma and neuroblastoma cells show minimal peptide internalisation.
- The phagocytosis of labelled Amyloid β which was aggregated for 0, 2 or 4 days was compared. Increasing the aggregation of the peptide changes the kinetic profile of uptake by BV-2 cells, and longer aggregation times resulted in greater engulfment over a 24h period.

Blockade of scavenger receptors inhibits engulfment of A β

