

# Rapid and Specific Detection of STEC Strains O157:H7, O26 and O111 Using a Label-Free Biosensor System



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## Introduction

### Pathogens

- Escherichia coli* O157:H7, a strain of enterohemorrhagic, Shiga toxin-producing *E. coli* (STEC), causes gastroenteritis, resulting in (bloody) diarrhea and sometimes acute kidney failure due to HUS (hemolytic-uremic syndrome).
- E. coli* O26 and O111 are STEC strains, similar to O157:H7, and are emerging threats to human health.
- Salmonella enterica* serovar Typhimurium causes gastroenteritis, resulting in diarrhea, fever and abdominal cramping.

### Incidence

- Annually in the U.S. there are >100,000 cases of STEC and >1.4 million non-typhoidal *Salmonella* infections.

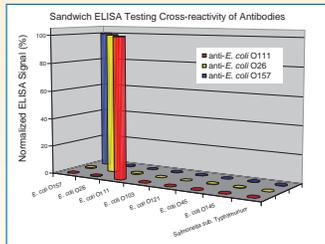
### Problem

- Detection of foodborne pathogens is the primary means of preventing contaminated foodstuffs from entering the market.
- Traditional immunoassays are the gold standard for identifying specific strains of bacteria. However, while specific, these immunoassays are time-consuming.
- Most current biosensor applications focus on specificity or sensitivity rather than assay speed.

## Materials and Methods

### Antibodies

- Polyclonal antibodies against *E. coli* O157 (KPL Cat. # 01-95-90), *E. coli* O26 (KPL Cat. # 01-95-92), *E. coli* O111 (KPL Cat. # 01-95-91) and *Salmonella* species (KPL Cat. # 01-91-99) were used as capture antibodies.



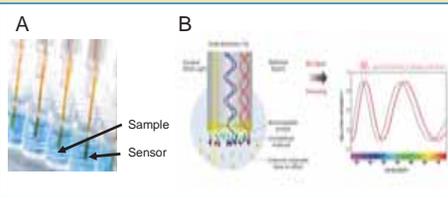
**Figure 1: Antibody specificity**  
Sandwich ELISA showing minimal cross-reactivity between anti-*E. coli* O111, O26 and O157.

- Bacteria were serially diluted in EDTA buffer (10 mM phosphate, 150 mM NaCl, 10 mM EDTA).
- Bacteria enumeration was accomplished by plating replicates on TSB agar.
- Bacteria were disrupted by sonication and 1% Triton X-100 prior to pelleting; supernatants were assayed.
- Human serum was diluted 1:10 in EDTA buffer before use.
- Hamburger (25 g/225 mL buffer) was sonicated, pelleted and filter sterilized before use.

## Aim

To develop methods using a commercially available biosensor in concert with anti-bacterial antibodies to expedite detection of foodborne pathogens.

## Biosensor



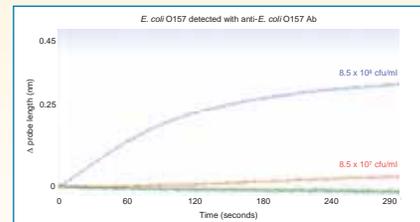
**Figure 2: Bi-layer Interferometry**  
(A) Fiber-optic biosensor tips (~1900  $\mu\text{m}^2$ ) in solution. (B) Model depicting principle of bi-layer interferometry.

- Bi-layer interferometry (BLI) is a label-free immunoassay, in which the interference pattern of white light reflected from two surfaces, an internal reference layer and a layer of immobilized antigen on the tip surface, is analyzed.
- Automated dip-and-read technology permits detection of binding events within 100-300 seconds.
- The spectral shift measured by the instrument is proportional to the thickness of the antibody + antigen on the end of the probe.

### Instrument

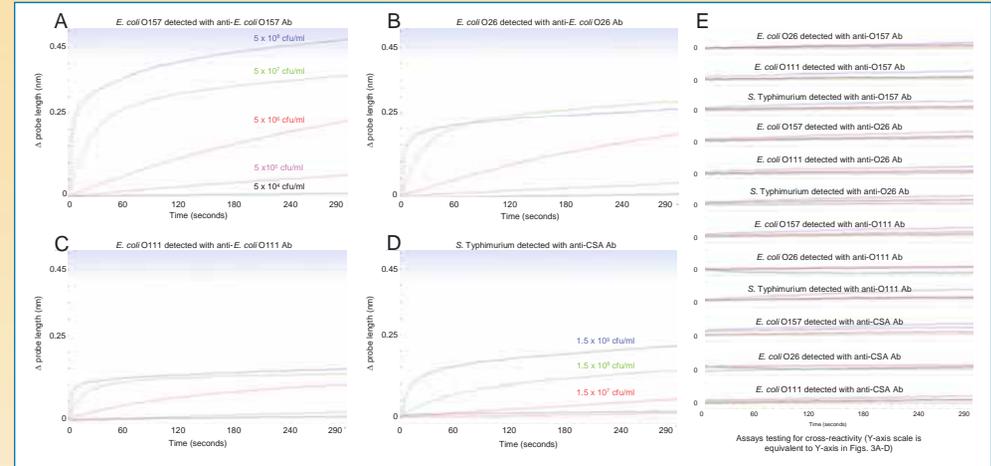
- Platform: ForteBio's Octet Red96.
- Biosensor: streptavidin functionalized.

## Results – Intact Bacteria



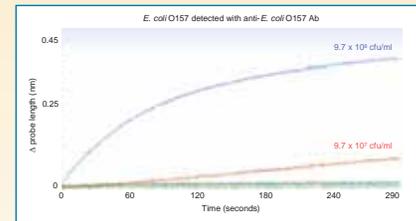
**Figure 3: Detection limits for intact bacteria**  
Detection of serial dilutions of *E. coli* O157 with cognate biotinylated polyclonal antibodies. Rapid detection of *E. coli* O157 was obtained within 300 seconds.

## Results – Disrupted Bacteria



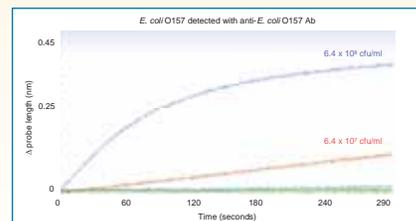
**Figure 4: Detection limits for disrupted bacteria**  
(A-D) Detection of sonicated and detergent treated bacteria with cognate biotinylated polyclonal antibodies. (E) Spectrographic traces of antibody/bacteria interaction determining cross-reactivity. Scale on parts A-E are the same, with the limit on Fig. 3E set at 0.05 nm. For all *E. coli* strains: Blue =  $5 \times 10^6$  cfu/ml, Green =  $5 \times 10^5$  cfu/ml, Red =  $5 \times 10^4$  cfu/ml, Pink =  $5 \times 10^3$  cfu/ml and Black =  $5 \times 10^2$  cfu/ml. For *Salmonella* Typhimurium experiments, Blue =  $1.5 \times 10^6$  cfu/ml, Green =  $1.5 \times 10^5$  cfu/ml, Red =  $1.5 \times 10^4$  cfu/ml, Pink =  $1.5 \times 10^3$  cfu/ml and Black =  $1.5 \times 10^2$  cfu/ml.

## Results – Beef Extract



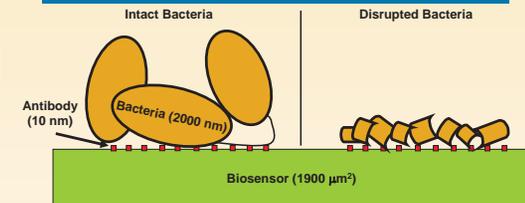
**Figure 5: Detection limits of *E. coli* O157 in hamburger extract**  
Macerated hamburger supernatant was diluted 1:10 in EDTA buffer. Detection of serial dilutions of *E. coli* O157 with cognate biotinylated polyclonal antibodies.

## Results – Human Serum



**Figure 6: Detection limits of *E. coli* O157 in human serum**  
Human serum was diluted 1:10 in EDTA buffer. Detection of serial dilutions of *E. coli* O157 with cognate biotinylated polyclonal antibodies.

## Model



**Figure 7: BLI Biosensor Model System**

The BLI biosensor is dependent on efficient packing for transmission of light. Small molecules bound to antibodies form a densely packed surface and are generally straightforward to detect. Intact bacteria, however, have asymmetrical densities, and almost certainly attach with a non-uniform packing order presenting challenges to detection without processing.

## Summary

- Detection of bacterial pathogens has been demonstrated in multiple matrices, for the first time, using the commercially available Octet Red96 biosensor from ForteBio.
- Bi-layer interferometry, in combination with KPL's polyclonal BacTrace® antibodies, is a viable method for rapid, specific detection of various bacteria strains, including the pathogens of *E. coli* O157, *E. coli* O26, *E. coli* O111 and *Salmonella* Typhimurium.
- Using BLI, as few as  $5 \times 10^5$  CFU/mL of bacteria can be detected in less than four hours (total assay time).
- Time consuming enrichment steps were avoided.
- Processing of the samples, in order to make a more uniform layer on the biosensor, was needed for higher levels of sensitivity.