

# SARTORIUS



## The Next Generation of SPR-Based Interaction Analysis

Octet® SF3 SPR

**A robust, high-throughput, low maintenance SPR solution**

- State-of-the-art microfluidics and optimized electronic designs
- Complete kinetics and affinity for up to 768 samples in a single unattended assay
- Eliminate the need to prepare multiple dilution series using OneStep® Injection Technology
- Determine full kinetics and affinity in the presence of multiple competitors

The Octet® SF3 SPR system from Sartorius employs a novel gradient injection technology that combines the accuracy and sensitivity of traditional SPR analysis. Based on the well-established concept of Taylor dispersion injections, Octet® SF3 SPR provides the benefit of simpler assay setups, shorter run times, reduced sample consumption, and higher throughput. Octet® SF3 precisely delivers sample analyte to the SPR flow cell in a continuous concentration gradient using an innovative sample dispersion and delivery system. A wide range of concentrations is covered in a single injection, eliminating the need to run multiple dilutions of sample to obtain accurate kinetic and affinity constants. OneStep® and NeXtStep™ gradient injections are two examples of how the Octet® SF3 can improve data quality and reduce the time to both develop and run assays.

## OneStep® Injections

A OneStep® gradient injection disperses analyte in the sample through an injection line filled with buffer in the Octet® SF3 fluidics en route to the SPR flow cell. This method, based on Taylor dispersion, produces a sigmoidal concentration gradient of analyte in the injection line. As the sample gradient flows over the sensor chip, real time, label-free binding data is collected in real time, incorporating the full range of analyte concentrations presented to the surface, from low to high. Figure 1 shows the injection line and the analyte concentration at the SPR flow cell.

## Improve Kinetic Characterization

Traditional SPR kinetic characterization relies on analyzing the time-resolved binding of an interaction over multiple concentrations of analyte. OneStep® gradient injections produces different analyte concentrations over time, from a single injection – eliminating the need for a dilution series of analyte and requiring only one sample, one concentration, and one injection to characterize the kinetics of an affinity interaction. This:

- Saves time and sample material
- Reduces the variance introduced by pipetting multiple samples and target immobilization among different channels
- Improves analysis of unstable targets which need to be tested quickly before all activity is lost

A OneStep® Injection's high-resolution concentration gradient also enables analyses as accurate as multi-cycle kinetics used in traditional SPR. OneStep® Gradient Injections diffuse a single concentration of analyte into a moving stream of buffer to create an analyte concentration gradient of at least 3 orders of magnitude, encompassing thousands of concentration data points. This heightened resolution is especially important for complex or heterogeneous interactions and is helpful in identifying non-specific or promiscuous interactions.

Analyte diffusion coefficients can also be determined from a OneStep® analysis. Diffusion can offer additional insight into the binding analysis, as the solution behavior of the analyte and formation of aggregates or higher order species can have an impact. This information is not accessible with traditional SPR measurements.

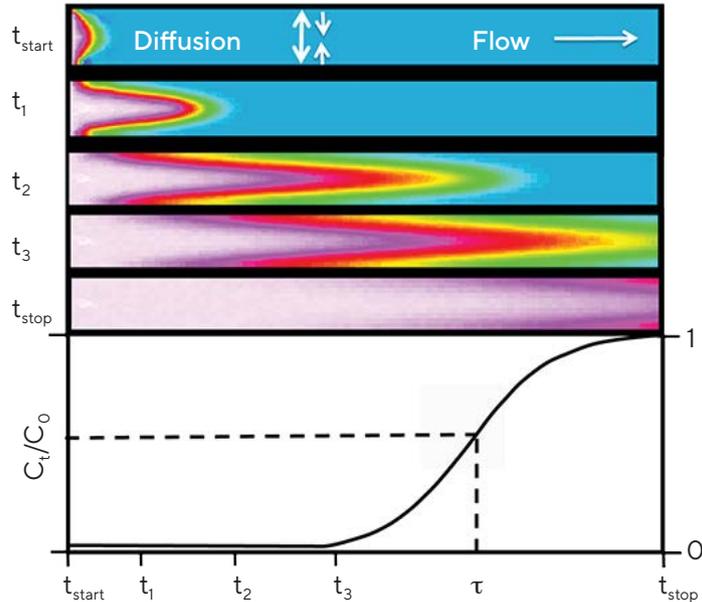


Figure 1: OneStep® gradient formation in the injection line (top), with the corresponding analyte concentration measured within the flow cell (bottom). Blue color indicates the running buffer and pink color indicates the analyte. The gradient formation and its relationship to analyte concentration at the flow cell is illustrated using five simulated snapshots ( $t_{\text{start}} - t_{\text{stop}}$ ) of the injection line at different times, and shows that a single injection can be used to assess a full analyte concentration series.

## Reduce Assay Development Time

Traditional SPR assay development is cumbersome and begins with finding a suitable method to immobilize the target molecule and then finding optimal conditions to observe binding of the analyte. An analyte binding test is often performed to determine activity of the target and to catch a glimpse of the binding kinetics between analyte and target.

OneStep® Injections simplify assay development by performing a full kinetic characterization at the time of the binding test. When a mid to high concentration of analyte is analyzed with a OneStep® Injection, a concentration gradient is produced that allows for the identification of optimal conditions as well as characterization of the interaction under the present conditions. Even if the maximum concentration used on the Octet® SF3 is high enough to saturate the binding interaction, the OneStep® gradient typically introduces low enough concentrations to accurately determine binding kinetics and affinity.

Time consuming linear DMSO calibration curves corrections can also be alleviated. Traditional SPR requires at least six DMSO standards to produce a linear calibration curve. The refractive index correction to account for varying concentrations of DMSO in samples can be performed with OneStep® micro-calibration injections on the Octet® SF3 system with just two DMSO standards. It's also more amenable to repeat calibrations to correct for changing sensor compositions (decaying protein surface, build-up of non-specific analytes, etc.).

## Work with Difficult Samples

### Unstable or unregenerable targets.

The immobilization of biomolecules on SPR sensor chips is a common pain point, as it can lead to destabilized molecules or the requirement to regenerate the immobilized molecule multiple times after analyte binding using standard multi-cycle kinetics. A OneStep® Injection's more rapid measurement can accommodate rapidly decaying immobilized molecule activity, and unregenerable molecules benefit since it does not require multiple analyte injections and regeneration of the immobilized molecule.

**Complex interactions.** Heterogeneity either on the part of the analyte or the immobilized target can be difficult to resolve in SPR assays. The heterogeneity observed in binding response curves that display the simultaneous binding of more than one event – e.g. two different forms of analyte to one target, one analyte to two different forms of target, etc. When assay development does not eliminate the complexity, deconvoluting these multiple events with traditional SPR requires testing up to 10 analyte concentrations to accurately determine more than one set of kinetic parameters ( $k_a$ ,  $k_d$ ,  $R_{max}$ ). A single OneStep® injection has the analytical resolution to deconvolute and analyze multiple kinetic parameter sets because the analyte concentration is a function of time, providing thousands of analyte concentrations in one gradient injection.

## Analyze a Wider Range of Molecules

The molecular weight of analytes is not limited with the OneStep® method. It has been successfully used to analyze interactions of small molecule fragments ( $\geq 70$  Da), small molecule compounds (100–400 Da), DNA/RNA oligonucleotides, lipids, peptides, proteins, protein oligomers, and aggregate species.

### Kinetics and Affinity Data from Primary Screens

In screening fragments, OneStep® Injections have been demonstrated to obtain reliable kinetics ( $k_a$ ,  $k_d$ ) and affinity ( $K_D$ ) data directly from the primary screen, combining the first three steps (initial screen, primary yes/no screen, affinity  $K_D$  screen) in the traditional SPR workflow into one step. The Octet® SF3 can process a complete screen from a new library to characterized hits in less than 1 week, compared to 2.5 weeks for a traditional 4-channel SPR system. Identification of promiscuous binders is also easier with OneStep® Injections, as the gradient resolves linear and super-stoichiometric binding events.



# NeXtStep™ Injections

NeXtStep™ Injections are another type of gradient injection where two samples are dispersed with one another, producing a crossed (as sample B concentration decreases, sample A concentration increases respectively) sigmoidal concentration profile (Figure 2). This is distinct from OneStep® Injections, which disperse one analyte sample with buffer. The two different samples injected by NeXtStep™ can then be used for competition and inhibition gradient assays.

## Increase Speed for Competition/Inhibition Assays

When screening for competition and inhibition, NeXtStep™ Injections provide increased speed and decreased sample consumption compared to traditional SPR assays. NeXtStep™ Injections require only one sample per competition/inhibition analyte (compared to eight samples with traditional SPR) and is nearly five-fold faster per analyte than a comparable SPR assay. NeXtStep™ methods employ multiple assay formats to

determine competition mechanisms or inhibition concentrations ( $IC_{50}$ ) in a single injection. There is no need for a sample dilution series as the NeXtStep™ gradient measures a wide concentration range encompassing all concentrations necessary for analysis.

## Reliable Data

Taylor Dispersion is a well-established effect in fluid mechanics that has been accepted for over half a century. The OneStep method and application was first published in 2012<sup>1</sup>, and since then, users of the technology have published their results in numerous peer-reviewed journals.

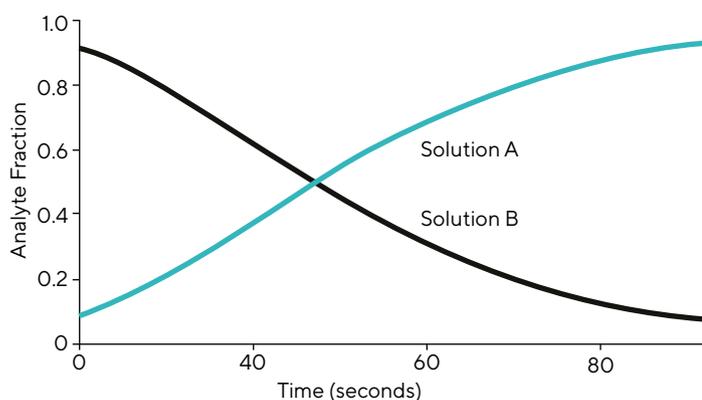


Figure 2: Time course charting the fraction of each solution as it passes through the flow cell. At the beginning of the injection, the solution almost entirely consists of solution B (the competitor molecule), however as the injection progresses, it is gradually replaced by solution A (the analyte + competitor molecule), until the final stages of the injection, which consists primarily of solution A.

## References

1. Modeling Taylor Dispersion Injections: Determination of Kinetic/Affinity Interaction Constants and Diffusion Coefficients in Label-Free Biosensing, Quinn JG, Anal. Biochem., 421(2) 391-410, 2012.

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